

HIV-2 Infection

This is a PDF version of the following document:

Module 6: [Key Populations](#)

Lesson 8: [HIV-2 Infection](#)

You can always find the most up-to-date version of this document at
<https://www.hiv.uw.edu/go/key-populations/hiv-2/core-concept/all>.

Background

Comparison of HIV-1 and HIV-2

Human immunodeficiency virus (HIV) is categorized into two main types: HIV-1 and HIV-2. Although HIV-1 and HIV-2 have the same routes of transmission, and both can cause acquired immunodeficiency syndrome (AIDS), important differences exist between the viruses in terms of epidemiology, natural history, diagnosis, and management.[1,2] Compared with individuals with HIV-1, persons with HIV-2 typically have attenuated clinical progression and lower rates of sexual and perinatal HIV transmission.[3,4,5,6] Significant differences exist in the antiretroviral management of HIV-1 and HIV-2.[2] Clinicians should become familiar with the differences between HIV-1 and HIV-2 and maintain a high index of suspicion of HIV-2 in persons from regions where HIV-2 circulates. In addition, it is important to understand the same individual can acquire both HIV-1 and HIV-2.[7]

HIV-2 Epidemiology

HIV-2 Globally

The World Health Organization (WHO) estimated there were 39 million people living with HIV worldwide at the end of 2022, but current HIV-2 specific prevalence estimates are lacking.[8] Since the beginning of the HIV pandemic, approximately 1 to 2 million people have been infected with HIV-2, including those with both HIV-1 and HIV-2.[1,9,10,11] Most persons infected with HIV-2 reside in West Africa, or in countries that have strong colonial or socioeconomic ties with West Africa, most notably France, Spain, and Portugal; HIV-2 has been reported in significant numbers in several former Portuguese colonies, including Angola, Mozambique, Brazil, and India (mainly in the states of Goa and Maharashtra and to a lesser degree in some southern regions).[1] During approximately the last two decades, HIV-2 prevalence has declined in several West African countries, but the reasons for this are unclear.[12]

HIV-2 in the United States

In the United States, it is estimated that fewer than 1% of persons with HIV have HIV-2. The first case of HIV-2 in the United States was reported in 1987.[13] The vast majority of persons diagnosed with HIV-2 in the United States have emigrated from an HIV-2 endemic region or had exposure to a person from an HIV-2 endemic region.[14] In addition, most of the earlier reported small number of cases of HIV-2 in the United States have been clustered in the northeast.[14,15] More recently, the Centers for Disease Control and Prevention (CDC) analyzed data from the National HIV Surveillance System (NHSS) that included data on persons diagnosed with HIV-1, HIV-2, or both during 2010–2017 in the United States and 6 dependent areas.[16] Overall, this report concluded the diagnosis of HIV-2 was rare during this time period.[16] The image series below summarizes the major characteristics of persons in the report who were diagnosed with HIV-2 (Figure 1).[16]

Pathogenesis, Transmission, and Natural History of HIV-2

Pathogenesis of HIV-2

Relative to HIV-1, HIV-2 is less virulent and is characterized by lower plasma HIV RNA levels, a slower decline in CD4 cell counts, and a longer time to progress to AIDS.[2,12,17] For example, in West Africa, investigators prospectively followed 133 persons with HIV-2 who were not receiving antiretroviral therapy from 1991 through 2009 and found HIV-2 RNA levels remained consistently low, with 36 to 42% of the HIV-2 RNA levels falling in the range of less than 100 copies/mL.[18] This same study showed the rate of disease progression and mortality rates correlated with the baseline plasma HIV-2 RNA levels obtained in 1991: individuals with baseline HIV-2 RNA levels less than 100 copies/mL had very low mortality.[18] In another longitudinal follow-up study in West Africa, investigators have shown that HIV-2 is more pathogenic than previously thought and that most people with untreated HIV-2 will eventually develop AIDS within 15 years of HIV acquisition (Figure 2).[19] The exact reasons for the differences in pathogenicity of HIV-1 and HIV-2 remain incompletely defined, but both intrinsic viral factors and innate and adaptive immunity are likely to play important roles.[17] Persons with HIV-2 who suffer immunologic decline develop similar opportunistic infections as seen in individuals with HIV-1.[17]

Transmission of HIV-2

In natural history perinatal studies conducted in West Africa (in the pre-antiretroviral era), the rate of perinatal transmission of HIV-2 was significantly lower than for HIV-1, typically less than 5% versus approximately 25%.[6,20] Sexual transmission and genital shedding is also less efficient in persons with HIV-2.[5,21,22,23] Acquisition of HIV-2 does not provide protection against infection with HIV-1.[24]

Effect of HIV-2 Coinfection on HIV-1 Progression

Limited data exist on the natural disease progression in persons with concomitant HIV-1 and HIV-2 infection; in West Africa, up to 15% (approximately) of individuals have dual infection, though the proportion of persons with HIV-2 mono-infection has decreased as the prevalence of HIV-2 has decreased.[17,25] Unfortunately, initial infection with HIV-2 does not appear to protect against subsequent HIV-1 acquisition, as was initially reported in a cohort of female sex workers in Dakar, Senegal.[24,26,27] One study suggested that persons with both HIV-1 and HIV-2 have slower disease progression and delayed death when compared with those who have HIV-1 alone, with the greatest benefit occurring when infection with HIV-2 precedes HIV-1 infection.[25] Other studies, including a robust meta-analysis, found no survival benefit in persons dually infected with HIV-1 and HIV-2 when compared with persons who have HIV-1 alone.[26,28]

2014 Surveillance Case Definition for HIV-2 Infection

In 2014, the Centers for Disease Control and Prevention released a Revised Surveillance Case Definition for HIV Infection that added specific criteria for defining a case of HIV-2, which was not part of the 2008 case definition.[\[29\]](#) To classify an adult as having HIV-2, one or more of the following laboratory criteria are necessary:

- FDA-approved HIV-1/HIV-2 type-differentiating antibody test result positive for HIV-2 and negative for HIV-1,
- Positive HIV-2 Western blot result and negative or indeterminate HIV-1 Western blot result,
- Positive qualitative HIV-2 nucleic acid test (NAT),
- Detectable quantitative HIV-2 NAT (viral load), *or*
- Laboratory results are interpreted as consistent with HIV-2 infection by a laboratory expert experienced in differentiating HIV-2 from HIV-1 (if laboratory evidence for HIV-2 is ambiguous).

In addition, the 2014 Revised Surveillance Case Definition for HIV Infection classifies an individual as having dual infection (with HIV-1 and HIV-2) if both an HIV-1 NAT and an HIV-2 NAT are positive.[\[29\]](#)

Diagnostic Testing for HIV-2

Approach to Diagnostic Testing for HIV-2

In CDC screening guidelines for HIV issued prior to 2014, specific HIV-2 testing was recommended only for persons with known HIV-2 risk factors.[30] With this older HIV testing algorithm, which utilized HIV enzyme immunoassay (EIA) as the screening test and HIV-1 Western blot as the confirmatory test, the diagnosis of HIV-2 was often missed because HIV EIA testing detects both HIV-1 and HIV-2, but does not distinguish between them; in addition, the traditional HIV-1 Western blot fails to detect HIV-2 (or it indicates an indeterminate or false positive result for HIV-1).[14,30,31] It is important to note that HIV-1 RNA and DNA assays do not reliably detect HIV-2.[1,31] Using the tests as recommended in the CDC and Association of Public Health Laboratories (APHL) HIV diagnostic algorithm can reliably detect HIV-2; the HIV-1/HIV-2 antigen-antibody test is the recommended initial test, followed by an HIV-1/HIV-2 differentiation immunoassay if the initial test is positive (Figure 3).[31,32] In some circumstances, such as an indeterminate HIV-2 test on the HIV-1/HIV-2 differentiation immunoassay, HIV-2 NAT diagnostic testing is recommended.[33] Diagnostic HIV-2 qualitative testing is now available for diagnostic purposes through two laboratories: the University of Washington Laboratory Medicine (HIV-2 DNA/RNA Qualitative) and the New York State Department of Health (HIV-2 Qualitative RNA Detection). It is important to recognize that about 30 to 40% of persons with HIV-2 who are not receiving antiretroviral therapy have undetectable plasma HIV-2 RNA levels; for this reason, HIV-2 RNA testing alone is not a reliable diagnostic test.[34,35,36]

Interpretation of Diagnostic Tests in HIV-2

- **Enzyme Immunoassay (EIA):** A person with HIV-2 will likely have a positive HIV enzyme immunoassay (EIA) regardless of which test is used because most, but not all, EIA tests detect both HIV-1 and HIV-2.[30] Commercially available EIAs do not generally differentiate between HIV-1 and HIV-2, although there is a specific HIV-2 EIA that is FDA-approved (Genetic Systems HIV-2 EIA).
- **Point-of-Care Tests:** Several point-of-care HIV tests are FDA-approved for the detection of HIV-2, including the OraQuick Advance Rapid HIV-1/2 Antibody Test, Clearview HIV 1/2 STAT-PAK, Clearview COMPLETE HIV 1/2, INSTI HIV-1/HIV-2 Rapid Antibody Test, Architect HIV Ag/Ab Combo Assay, and Alere Determine HIV-1/2 Ag/Ab. These point-of-care tests do not distinguish HIV-1 from HIV-2 infection.
- **Differentiation Assays for HIV-1/HIV-2:** In the United States, the Geenius HIV 1/2 Supplemental Assay, and the BioPlex 2200 HIV Ag-Ab are FDA-approved for differentiating HIV-1 from HIV-2 infection.[37,38,39] The Geenius test can detect four antibodies to HIV-1 (p31, gp160, p24, and gp41) and two antibodies to HIV-2 (gp36 and gp140) (Figure 4).[37,40] The CDC recommended algorithm for HIV testing utilizes the HIV-1/HIV-2 differentiation assay as the second step in the algorithm.[31] The CDC has issued a technical update on HIV-1/2 differentiation assays that provides guidance for three results that may occur in the Geenius assay that were not previously seen with the Multispot HIV differentiation assay: HIV-2 positive with HIV-1 cross reactivity, HIV-2 indeterminate, and HIV indeterminate.[33] If the result is HIV-2 positive with HIV-1 cross reactivity, the CDC recommends considering this result as positive for HIV-2 infection. For specimens with either HIV-2 indeterminate or HIV indeterminate results, additional testing is required (often including HIV-2 nucleic acid testing), and guidance in the technical update should be followed carefully; ideally, expert consultation is obtained in this setting.
- **HIV Western Blot:** The HIV Western blot is no longer routinely used for HIV diagnosis and is not recommended in the CDC HIV testing algorithm. Infection with HIV-2 may cause a negative, indeterminate, or positive HIV-1 Western blot due to cross-reacting antibodies. Persons with HIV-2 often have an indeterminate HIV-1 Western blot pattern, with the presence of Gag bands (p55, p24, or p17) and Pol bands (p66, p51, or p31), but the absence of Env bands (gp160, gp120, or gp41) (Figure 5).[1,15] The Western blot pattern in persons with HIV-2 occurs because HIV-1 and HIV-2 have 60% similarity in the regions encoding *gag* and *pol*, compared with only 30 to 40% similarity in the region encoding *env*.[41] In the United States, HIV-2 Western blot tests are not FDA-approved, but some HIV-2 supplemental HIV-2 antibody tests are commercially available through reference laboratories.[1]

- **Qualitative Plasma HIV-2 RNA:** An HIV-2 nucleic acid test is typically utilized to confirm a positive HIV-2 differentiation assay or to provide additional information in the setting of an indeterminate result on the HIV Geenius differentiation assay. A positive qualitative HIV-2 RNA will confirm a diagnosis of HIV-2. A negative HIV-2 plasma RNA test, however, does not rule out HIV-2, since approximately 30 to 40% of persons with untreated chronic HIV-2 have undetectable plasma HIV-2 RNA levels.[[34](#),[35](#),[36](#)] To address this issue, investigators have developed a qualitative assay that detects HIV-2 total nucleic acid in patient peripheral blood mononuclear cells; this assay can detect HIV-2 DNA and RNA in persons with HIV-2 who have undetectable plasma HIV-2 RNA levels.[[42](#)]

Quantitative HIV-2 RNA Testing

The HIV-1 nucleic acid amplification tests (NAAT) do not reliably detect or quantitate HIV-2. Quantitative HIV-2 RNA viral load assays for monitoring response to therapy are available through the University of Washington Laboratory Medicine ([HIV-2 RNA Quantitation](#)) and the New York State Department of Health ([HIV-2 Quantification](#)).[2]

Antiretroviral Susceptibility and Resistance

The following summarizes what is known related to HIV-2 susceptibility and resistance to medications in specific antiretroviral medication classes.

Nucleoside Reverse Transcriptase Inhibitors (NRTIs)

In general, nucleoside reverse transcriptase inhibitors (NRTIs) are active against HIV-2, but due to naturally occurring polymorphisms in HIV-2, they may have less activity and a lower genetic barrier of resistance to HIV-2 than to HIV-1.[\[43\]](#) Although HIV-1 and HIV-2 share some classic NRTI resistance mutations, such as the M184V mutation, which causes high-level resistance to lamivudine and emtricitabine, HIV-2 often follows different resistance pathways than HIV-1. For example, HIV-2 resistance to zidovudine occurs through the Q151M mutation rather than through the common thymidine analog mutation (TAM) pathways typically observed with HIV-1.[\[44\]](#) Studies have produced conflicting results regarding the frequency of the K65R mutation in HIV-2 and its impact on susceptibility to tenofovir and abacavir.[\[43,45,46\]](#) Available data suggest the development of the Q151M mutation in combination with a K65R or M184V mutation results in resistance to zidovudine, lamivudine, and emtricitabine; the presence of all three mutations together causes broad NRTI class resistance.[\[47\]](#)

Non-Nucleoside Reverse Transcriptase Inhibitors (NNRTIs)

Multiple studies have shown that HIV-2 has intrinsic resistance to non-nucleoside reverse transcriptase inhibitor (NNRTI) drugs.[\[43,48\]](#) This intrinsic resistance occurs because the Y181I and Y188L substitutions are natural polymorphisms present in HIV-2 strains and these mutations alter the NNRTI binding pocket in the reverse transcriptase enzyme, rendering it less receptive to NNRTI medications; this includes the newest NNRTI, doravirine, as well as the long-acting injectable rilpivirine as part of the injectable cabotegravir and rilpivirine combination.[\[49\]](#)

Integrase Strand Transfer Inhibitors (INSTIs)

Accumulating evidence suggests that INSTIs usually have potent activity against HIV-2.[\[2,50,51,52\]](#) Dolutegravir, bictegravir, and cabotegravir have in vitro activity against HIV-2.[\[53,54,55\]](#) A recent, small, single-arm, clinical trial from Portugal showed that dolutegravir, when paired with 2 NRTIs, was effective for treatment-naïve persons with HIV-2 and another small study showed modest efficacy with dolutegravir in persons with HIV-2 who had resistance to first-generation INSTIs.[\[56,57\]](#) Elvitegravir has activity against HIV-2 and, as in HIV-1, there is a high degree of cross-resistance between raltegravir and elvitegravir.[\[50\]](#) As seen with HIV-1, raltegravir and elvitegravir have a relatively low barrier to resistance to HIV-2, perhaps even lower with HIV-2 than with HIV-1 due to intrinsic polymorphisms at secondary integrase sites.[\[56,58\]](#) The resistance that affects the susceptibility of HIV-2 to INSTIs shares some similarities with those of HIV-1, though some key differences exist.[\[54,59,60\]](#) For example, the integrase mutations N155H or Q148R confer resistance to raltegravir in both HIV-2 and HIV-1, but for HIV-2, the Y143 pathway requires secondary mutations to cause significant resistance (contrary to what is observed with HIV-1).[\[59\]](#) A study revealed a new resistance pathway of HIV-2 to INSTIs that involves a 5-amino acid insertion at codon 231 of the HIV-2 integrase, which is a region in the integrase C-terminal domain.[\[61\]](#) This insertion mutation results in high-level resistance to elvitegravir and raltegravir and moderate-level resistance to dolutegravir.[\[61\]](#) Resistance to bictegravir was observed in several isolates, but overall, it retained the most potent INSTI in isolates with the codon 231 insertion mutation (231INS).[\[61\]](#)

Protease Inhibitors

Several studies have reported that HIV-2 has inherent partial or full resistance to some protease inhibitors, with only lopinavir-ritonavir, darunavir, and saquinavir having clinically useful activity against HIV-2.[\[43,62,63,64,65\]](#) Investigators have identified four residues at amino acid positions—32, 47, 76, and

82—in the protease binding cleft that differ between HIV-2 and HIV-1 and predict protease inhibitor sensitivity; changes at these four amino acids can confer class-wide resistance to PIs.[66] Studies have shown that fewer HIV protease mutations are necessary for resistance to develop in HIV-2 compared with HIV-1. Due to innate polymorphisms, a lower number of mutations are required for HIV-2 to become resistant to PIs as compared to HIV-1. Similar to treatment for HIV-1, if a protease inhibitor is used to treat HIV-2, boosting with ritonavir or cobicistat is recommended. Based on available data, the recommended protease inhibitor-based regimens for HIV-2 consist of boosted darunavir, lopinavir, or saquinavir, in combination with two NRTIs.[2] Notably, atazanavir is not recommended for the treatment of HIV-2.

Entry Inhibitors

- **Attachment Inhibitors:** Limited data suggest that temsavir, the active moiety of fostemsavir, is not active against HIV-2 in vitro.[67,68]
- **Post-Attachment Inhibitors:** Limited in vitro data shows that ibalizumab is active against HIV-2.[69]
- **CCR5 Inhibitor:** The efficacy of the CCR5 inhibitor, maraviroc, is uncertain since HIV-2 can use several different co-receptors to enter cells, and there is no commercial HIV-2 co-receptor tropism screening assay that would determine whether the patient has pure CCR5-tropic HIV-2.[1,2,70,71]
- **Fusion Inhibitors:** In vitro data have shown HIV-2 has intrinsic resistance to the fusion inhibitor, enfuvirtide.[48] Resistance to enfuvirtide correlates with genetic diversity at the target regions for the drug, namely the HR1 domain of the viral gp41 region.[72]

Capsid Inhibitors

Limited in vitro data shows that lenacapavir, currently the only available HIV capsid inhibitor, has activity against HIV-1 and HIV-2, but this activity is 11- to 25-fold less than against HIV-1.[68,73] More data are needed on lenacapavir and HIV-2.

HIV-2 Treatment Studies

Clinical Trials for the Treatment of HIV-2

Limited data from clinical trials that inform guidance on the optimal timing or regimen for initial antiretroviral therapy in persons with HIV-2, which include three small, single-arm trials of INSTI-based (raltegravir, elvitegravir, and dolutegravir) treatment for initial antiretroviral therapy.[\[4,57,74\]](#) In addition, a randomized controlled trial comparing tenofovir DF-emtricitabine plus raltegravir versus tenofovir DF-emtricitabine plus lopinavir-ritonavir for the treatment of persons with HIV-2 has been completed, but the results have not yet been reported. There are no clinical data on the use of bictegravir-tenofovir alafenamide-emtricitabine for the treatment of HIV-2, but in vitro data suggest that HIV-2 is highly sensitive to bictegravir.[\[53\]](#) The lack of data on HIV-2 treatment is due to a combination of factors, including the low prevalence of HIV-2 (especially in the United States and Europe) and the lower virulence of HIV-2 compared with HIV-1, which has made investigation of antiretroviral therapy for HIV-2 less of a priority than for those with HIV-1.[\[1\]](#)

HIV-2 Treatment Recommendations

Timing of Initiating Antiretroviral Therapy with HIV-2

Although individuals with HIV-2 generally have a slower disease progression than persons with HIV-1, they generally have a less robust CD4 count increase in response to antiretroviral treatment.[\[75,76,77,78\]](#) This poor CD4 cell count response to antiretroviral therapy in persons with HIV-2 suggests that persons with HIV-2 should start antiretroviral therapy without delay. In addition, early treatment of HIV-2, in theory, would reduce transmission of HIV-2 to others.

- **Recommendation:** The Adult and Adolescent ARV Guidelines recommend starting antiretroviral therapy at or soon after HIV-2 diagnosis to prevent disease progression and transmission of HIV-2 to others.[\[2\]](#)

Recommended Antiretroviral Regimens for Treatment of HIV-2

The following summarizes key recommendations in the Adult and Adolescent ARV Guidelines for the treatment of HIV-2 (without HIV-1 infection).[\[2\]](#)

- Based on experience with the treatment of HIV-1, a three-drug antiretroviral regimen should be used to treat HIV-2 in order to maintain viral suppression and to avoid the development of resistance from suboptimal therapy.
- Since resistance testing is not commercially available for HIV-2, baseline resistance testing is not an option to guide initial therapy. Transmitted HIV-2 drug resistance has been reported, but to date, appears to be rare.[\[79,80\]](#)
- The preferred treatment regimen for persons with HIV-2 should consist of 2 NRTIs in combination with an INSTI (bictegravir, dolutegravir, elvitegravir, or raltegravir).
- The alternative regimen is two NRTIs plus a boosted protease inhibitor (darunavir or lopinavir) active against HIV-2. If a protease inhibitor is used, boosted darunavir may be preferred over boosted lopinavir because it is better tolerated.
- The following medications should not be used to treat HIV-2: any medication in the NNRTI class, fostemsavir, enfuvirtide, and long-acting injectable cabotegravir and rilpivirine.
- Persons with HIV-2 and hepatitis B virus (HBV) coinfection require an antiretroviral regimen that contains drugs with activity against both HIV-2 and HBV.
- For persons with multidrug-resistant HIV-2, ibalizumab and lenacapavir could be considered as part of a salvage regimen, based on in vitro activity.

Treatment of Persons with HIV-1 and HIV-2 Dual Infection

The following summarizes key recommendations in the Adult and Adolescent ARV Guidelines for the treatment of HIV-1 and HIV-2 dual infection.[\[2\]](#)

- Individuals with HIV-1 and HIV-2 dual infection should undergo baseline genotypic resistance testing for HIV-1; resistance testing for HIV-2 is not commercially available in the United States.[\[2,81\]](#) In this setting, if possible, monitoring of both HIV-1 and HIV-2 plasma RNA levels should be performed.[\[82,83\]](#)
- The preferred treatment of persons with HIV-1 and HIV-2 dual infection is 2 NRTIs in combination with an INSTI, even if the baseline HIV-2 plasma viral load is low or undetectable.
- All the preferred regimens for the treatment of HIV-1 have good activity against both HIV-1 and HIV-2.
- The following medications should not be used to treat persons with dual HIV-1 and HIV-2 infection: any medication in the NNRTI class, fostemsavir, enfuvirtide, and long-acting injectable cabotegravir and rilpivirine.
- Persons with HIV-1 and HIV-2 dual infection and HBV coinfection require an antiretroviral regimen that

contains drugs with activity against HIV-1, HIV-2, and HBV.

- For persons with multidrug-resistant HIV-1 and HIV-2, ibalizumab and lenacapavir could be considered as part of a salvage regimen, based on in vitro activity.

Clinical follow-up and Laboratory Monitoring for HIV-2

Until further HIV-2 treatment data are available, clinicians should follow recommendations for HIV-1 clinical management and HIV primary care, including opportunistic infection prophylaxis and laboratory monitoring on antiretroviral therapy.[2] The HIV-1 nucleic acid tests (NAT) do not reliably detect or quantitate HIV-2. Traditionally, because of the limited availability of HIV-2 RNA assays, response to antiretroviral therapy in most individuals with HIV-2 has been gauged only by regular clinical monitoring and repeated CD4 cell count monitoring. Quantitative HIV-2 RNA viral load assays for monitoring response to therapy are now available through the University of Washington Laboratory Medicine ([HIV-2 RNA Quantitation](#)) and the New York State Department of Health ([HIV-2 Quantification](#)).[2] Since several laboratories are now capable of performing quantitative HIV-2 RNA levels, the management of persons with HIV-2 on antiretroviral therapy should include routine monitoring of quantitative HIV-2 levels, similar to what is done for persons with HIV-1. Ongoing CD4 count monitoring is recommended in persons with HIV-2, even if their viral load is undetectable or suppressed.

Summary Points

- Compared with HIV-1, HIV-2 is a less virulent and less transmissible virus. More recent data has shown that HIV-2 is more pathogenic than previously thought, and most persons with HIV-2 will develop AIDS within 15 years.
- Infection with HIV-2 should be considered in persons with risk factors for HIV-2 acquisition and in persons with a clinical illness (such as an AIDS-associated opportunistic infection) that suggests HIV infection but in whom testing for HIV-1 is negative.
- Use of the CDC and APHL HIV diagnostic testing algorithm (1) detects HIV-1 and HIV-2 infection in the initial screening test and (2) distinguishes HIV-1 and HIV-2 with the HIV differentiation assay used in the second step. A positive HIV-2 qualitative RNA or DNA confirms infection, but HIV-2 RNA alone is not reliable to rule out infection since approximately 30 to 40% of persons with HIV-2 have undetectable HIV-2 RNA levels.
- Persons with HIV-2 have poorer CD4 cell count responses to antiretroviral treatment relative to persons with HIV-1, so early diagnosis and early initiation of therapy for HIV-2 should be emphasized.
- HIV-2 has intrinsic resistance to all NNRTI drugs, some PIs, fostemsavir, and enfuvirtide.
- HIV-2 is generally susceptible to NRTIs, INSTIs, and certain PIs, although naturally occurring polymorphisms may result in a lower genetic barrier to resistance to HIV-2 than to HIV-1. Among the available PIs, lopinavir and darunavir have the best activity against HIV-2.
- The preferred antiretroviral treatment for HIV-2 infection is 2 NRTIs plus an INSTI. The alternative regimen is 2 NRTIs plus a boosted PI (darunavir or lopinavir).
- Individuals with HIV-1 and HIV-2 coinfection should receive an antiretroviral regimen that can effectively treat both viruses, ideally two NRTIs plus an INSTI.
- In the event of clinical or virologic failure on antiretroviral therapy, consultation with an expert in HIV-2 management is recommended.
- Laboratory diagnostics for HIV-2 are available through two laboratories: the University of Washington Laboratory Medicine (HIV-2 DNA/RNA Qualitative and RNA Quantitative) and the New York State Department of Health (HIV-2 RNA Qualitative and Quantitative).

Citations

1. Campbell-Yesufu OT, Gandhi RT. Update on human immunodeficiency virus (HIV)-2 Infection. Clin Infect Dis. 2011;52:780-7.
[\[PubMed Abstract\]](#) -
2. Panel on Antiretroviral Guidelines for Adults and Adolescents. Guidelines for the use of antiretroviral agents in adults and adolescents with HIV. Department of Health and Human Services. Considerations for antiretroviral use in special patient populations: HIV-2 infection. December 6, 2023.
[\[HIV.gov\]](#) -
3. Marlink R, Kanki P, Thior I, et al. Reduced rate of disease development after HIV-2 infection as compared to HIV-1. Science. 1994;265:1587-90.
[\[PubMed Abstract\]](#) -
4. Matheron S, Pueyo S, Damond F, et al. Factors associated with clinical progression in HIV-2 infected-patients: the French ANRS cohort. AIDS. 2003;17:2593-601.
[\[PubMed Abstract\]](#) -
5. Kanki PJ, Travers KU, MBoup S, et al. Slower heterosexual spread of HIV-2 than HIV-1. Lancet. 1994;343:943-6.
[\[PubMed Abstract\]](#) -
6. Burgard M, Jasseron C, Matheron S, et al. Mother-to-child transmission of HIV-2 infection from 1986 to 2007 in the ANRS French Perinatal Cohort EPF-CO1. Clin Infect Dis. 2010;51:833-43.
[\[PubMed Abstract\]](#) -
7. Evans LA, Moreau J, Odehouri K, et al. Simultaneous isolation of HIV-1 and HIV-2 from an AIDS patient. Lancet. 1988;2:1389-91.
[\[PubMed Abstract\]](#) -
8. UNAIDS. Fact Sheet. 2025.
[\[UNAIDS\]](#) -
9. Ariën KK, Abraha A, Quiñones-Mateu ME, Kestens L, Vanham G, Arts EJ. The replicative fitness of primary human immunodeficiency virus type 1 (HIV-1) group M, HIV-1 group O, and HIV-2 isolates. J Virol. 2005;79:8979-90.
[\[PubMed Abstract\]](#) -
10. Gottlieb GS, Raugi DN, Smith RA. 90-90-90 for HIV-2? Ending the HIV-2 epidemic by enhancing care and clinical management of patients infected with HIV-2. Lancet HIV. 2018;5:e390-e399.
[\[PubMed Abstract\]](#) -
11. Williams A, Menon S, Crowe M, et al. Geographic and population distributions of HIV-1 and HIV-2 circulating subtypes: a systematic literature review and meta-analysis (2010-2021). J Infect Dis. 2023 Aug 18. Online ahead of print.
[\[PubMed Abstract\]](#) -
12. Gottlieb GS. Changing HIV epidemics: what HIV-2 can teach us about ending HIV-1. AIDS. 2013;27:135-7.
[\[PubMed Abstract\]](#) -
13. Centers for Disease Control (CDC). AIDS due to HIV-2 infection--New Jersey. MMWR Morb Mortal Wkly

Rep. 1988;37:33-5.

[\[PubMed Abstract\]](#) -

14. Centers for Disease Control and Prevention. HIV-2 Infection Surveillance--United States, 1987-2009. MMWR Morb Mortal Wkly Rep. 2011;60:985-8.
[\[PubMed Abstract\]](#) -
15. Torian LV, Eavey JJ, Punsalang AP, et al. HIV type 2 in New York City, 2000-2008. Clin Infect Dis. 2010;51:1334-42.
[\[PubMed Abstract\]](#) -
16. Peruski AH, Wesolowski LG, Delaney KP, et al. Trends in HIV-2 Diagnoses and Use of the HIV-1/HIV-2 Differentiation Test - United States, 2010-2017. MMWR Morb Mortal Wkly Rep. 2020;69:63-6.
[\[PubMed Abstract\]](#) -
17. Nyamweya S, Hegedus A, Jaye A, Rowland-Jones S, Flanagan KL, Macallan DC. Comparing HIV-1 and HIV-2 infection: Lessons for viral immunopathogenesis. Rev Med Virol. 2013;23:221-40.
[\[PubMed Abstract\]](#) -
18. van der Loeff MF, Larke N, Kaye S, et al. Undetectable plasma viral load predicts normal survival in HIV-2-infected people in a West African village. Retrovirology. 2010;7:46.
[\[PubMed Abstract\]](#) -
19. Esbjörnsson J, Månsson F, Kvist A, et al. Long-term follow-up of HIV-2-related AIDS and mortality in Guinea-Bissau: a prospective open cohort study. Lancet HIV. 2018;S2352-3018(18)30254-6.
[\[PubMed Abstract\]](#) -
20. Adjorlolo-Johnson G, De Cock KM, Ekpini E, et al. Prospective comparison of mother-to-child transmission of HIV-1 and HIV-2 in Abidjan, Ivory Coast. JAMA. 1994;272:462-6.
[\[PubMed Abstract\]](#) -
21. Gottlieb GS, Hawes SE, Agne HD, et al. Lower levels of HIV RNA in semen in HIV-2 compared with HIV-1 infection: implications for differences in transmission. AIDS. 2006;20:895-900.
[\[PubMed Abstract\]](#) -
22. Hawes SE, Sow PS, Stern JE, Critchlow CW, Gottlieb GS, Kiviat NB. Lower levels of HIV-2 than HIV-1 in the female genital tract: correlates and longitudinal assessment of viral shedding. AIDS. 2008;22:2517-25.
[\[PubMed Abstract\]](#) -
23. Simon F, Matheron S, Tamalet C, et al. Cellular and plasma viral load in patients infected with HIV-2. AIDS. 1993;7:1411-7.
[\[PubMed Abstract\]](#) -
24. Schim van der Loeff MF, Aaby P, Aryioshi K, et al. HIV-2 does not protect against HIV-1 infection in a rural community in Guinea-Bissau. AIDS. 2001;15:2303-10.
[\[PubMed Abstract\]](#) -
25. Esbjörnsson J, Månsson F, Kvist A, et al. Inhibition of HIV-1 disease progression by contemporaneous HIV-2 infection. N Engl J Med. 2012;367:224-32.
[\[PubMed Abstract\]](#) -
26. Prince PD, Matser A, van Tienen C, Whittle HC, Schim van der Loeff MF. Mortality rates in people dually infected with HIV-1/2 and those infected with either HIV-1 or HIV-2: a systematic review and meta-

analysis. AIDS. 2014;28:549-58.

[\[PubMed Abstract\]](#) -

27. Travers K, Mboup S, Marlink R, et al. Natural protection against HIV-1 infection provided by HIV-2. Science. 1995;268:1612-5.
[\[PubMed Abstract\]](#) -
28. Schim van der Loeff MF, Jaffar S, Aveika AA, et al. Mortality of HIV-1, HIV-2 and HIV-1/HIV-2 dually infected patients in a clinic-based cohort in The Gambia. AIDS. 2002;16:1775-83.
[\[PubMed Abstract\]](#) -
29. Centers for Disease Control and Prevention. Revised surveillance case definition for HIV infection--United States, 2014. MMWR Recomm Rep. 2014;63:1-10.
[\[PubMed Abstract\]](#) -
30. O'Brien TR, George JR, Epstein JS, Holmberg SD, Schochetman G. Testing for antibodies to human immunodeficiency virus type 2 in the United States. MMWR Recomm Rep. 1992;41:1-9.
[\[PubMed Abstract\]](#) -
31. Centers for Disease Control and Prevention and Association of Public Health Laboratories. Laboratory Testing for the Diagnosis of HIV Infection: Updated Recommendations. Published June 27, 2014.
[\[CDC\]](#) -
32. Centers for Disease Control and Prevention and Association of Public Health Laboratories. 2018 Quick reference guide: Recommended laboratory HIV testing algorithm for serum or plasma specimens. Published January 27, 2018.
[\[CDC\]](#) -
33. Centers for Disease Control and Prevention (CDC). Technical Update on HIV-1/2 Differentiation Assays. August 12, 2016.
[\[CDC\]](#) -
34. Chang M, Gottlieb GS, Dragavon JA, et al. Validation for clinical use of a novel HIV-2 plasma RNA viral load assay using the Abbott m2000 platform. J Clin Virol. 2012;55:128-33.
[\[PubMed Abstract\]](#) -
35. Ekouévi DK, Avettand-Fènoël V, Tchounga BK, et al. Plasma HIV-2 RNA According to CD4 Count Strata among HIV-2-Infected Adults in the IeDEA West Africa Collaboration. PLoS One. 2015;10:e0129886.
[\[PubMed Abstract\]](#) -
36. Gottlieb GS, Sow PS, Hawes SE, et al. Equal plasma viral loads predict a similar rate of CD4+ T cell decline in human immunodeficiency virus (HIV) type 1- and HIV-2-infected individuals from Senegal, West Africa. J Infect Dis. 2002;185:905-14.
[\[PubMed Abstract\]](#) -
37. Herssens N, Beelaert G, Fransen K. Discriminatory capacity between HIV-1 and HIV-2 of the new rapid confirmation assay Geenius. J Virol Methods. 2014;208:11-5.
[\[PubMed Abstract\]](#) -
38. Ramos EM, Harb S, Dragavon J, Coombs RW. Clinical performance of the Multispot HIV-1/HIV-2 rapid test to correctly differentiate HIV-2 from HIV-1 infection in screening algorithms using third and fourth generation assays and to identify cross reactivity with the HIV-1 Western Blot. J Clin Virol. 2013;58 Suppl 1:e104-7.
[\[PubMed Abstract\]](#) -

39. Salmona M, Delarue S, Delaugerre C, Simon F, Maylin S. Clinical evaluation of BioPlex 2200 HIV Ag-Ab, an automated screening method providing discrete detection of HIV-1 p24 antigen, HIV-1 antibody, and HIV-2 antibody. *J Clin Microbiol.* 2014;52:103-7.
[\[PubMed Abstract\]](#) -
40. Fernández McPhee C, Álvarez P, Prieto L, et al. HIV-1 infection using dried blood spots can be confirmed by Bio-Rad Geenius™ HIV 1/2 confirmatory assay. *J Clin Virol.* 2015;63:66-9.
[\[PubMed Abstract\]](#) -
41. Motomura K, Chen J, Hu WS. Genetic recombination between human immunodeficiency virus type 1 (HIV-1) and HIV-2, two distinct human lentiviruses. *J Virol.* 2008;82:1923-33.
[\[PubMed Abstract\]](#) -
42. Chang M, Wong AJ, Raugi DN, et al. Clinical validation of a novel diagnostic HIV-2 total nucleic acid qualitative assay using the Abbott m2000 platform: Implications for complementary HIV-2 nucleic acid testing for the CDC 4th generation HIV diagnostic testing algorithm. *J Clin Virol.* 2017;86:56-61.
[\[PubMed Abstract\]](#) -
43. Menéndez-Arias L, Alvarez M. Antiretroviral therapy and drug resistance in human immunodeficiency virus type 2 infection. *Antiviral Res.* 2014;102:70-86.
[\[PubMed Abstract\]](#) -
44. Boyer PL, Sarafianos SG, Clark PK, Arnold E, Hughes SH. Why do HIV-1 and HIV-2 use different pathways to develop AZT resistance? *PLoS Pathog.* 2006;2:e10.
[\[PubMed Abstract\]](#) -
45. Damond F, Matheron S, Peytavin G, et al. Selection of K65R mutation in HIV-2-infected patients receiving tenofovir-containing regimen. *Antivir Ther.* 2004;9:635-6.
[\[PubMed Abstract\]](#) -
46. Deuzing IP, Charpentier C, Wright DW, et al. Mutation V111I in HIV-2 reverse transcriptase increases the fitness of the nucleoside analogue-resistant K65R and Q151M viruses. *J Virol.* 2015;89:833-43.
[\[PubMed Abstract\]](#) -
47. Smith RA, Anderson DJ, Pyrak CL, Preston BD, Gottlieb GS. Antiretroviral drug resistance in HIV-2: three amino acid changes are sufficient for classwide nucleoside analogue resistance. *J Infect Dis.* 2009;199:1323-6.
[\[PubMed Abstract\]](#) -
48. Witvrouw M, Pannecouque C, Van Laethem K, Desmyter J, De Clercq E, Vandamme AM. Activity of non-nucleoside reverse transcriptase inhibitors against HIV-2 and SIV. *AIDS.* 1999;13:1477-83.
[\[PubMed Abstract\]](#) -
49. Ntemgwa ML, d'Aquin Toni T, Brenner BG, Camacho RJ, Wainberg MA. Antiretroviral drug resistance in human immunodeficiency virus type 2. *Antimicrob Agents Chemother.* 2009;53:3611-9.
[\[PubMed Abstract\]](#) -
50. Zheng Y, Lambert C, Arendt V, Seguin-Devaux C. Virological and immunological outcomes of elvitegravir-based regimen in a treatment-naïve HIV-2-infected patient. *AIDS.* 2014;28:2329-31.
[\[PubMed Abstract\]](#) -
51. Peterson K, Ruelle J, Vekemans M, Siegal FP, Deayton JR, Colebunders R. The role of raltegravir in the treatment of HIV-2 infections: evidence from a case series. *Antivir Ther.* 2012;17:1097-100.

[\[PubMed Abstract\]](#) -

52. Andreatta K, Miller MD, White KL. HIV-2 antiviral potency and selection of drug resistance mutations by the integrase strand transfer inhibitor elvitegravir and NRTIs emtricitabine and tenofovir in vitro. *J Acquir Immune Defic Syndr.* 2013;62:367-74.
[\[PubMed Abstract\]](#) -
53. Smith RA, Raugi DN, Wu VH, et al. Comparison of the Antiviral Activity of Bictegravir against HIV-1 and HIV-2 Isolates and Integrase Inhibitor-Resistant HIV-2 Mutants. *Antimicrob Agents Chemother.* 2019;63(5). pii: e00014-19.
[\[PubMed Abstract\]](#) -
54. Smith RA, Raugi DN, Pan C, et al. In vitro activity of dolutegravir against wild-type and integrase inhibitor-resistant HIV-2. *Retrovirology.* 2015;12:10.
[\[PubMed Abstract\]](#) -
55. Smith RA, Wu VH, Zavala CG, et al. In Vitro Antiviral Activity of Cabotegravir against HIV-2. *Antimicrob Agents Chemother.* 2018;62:e01299-18.
[\[PubMed Abstract\]](#) -
56. Descamps D, Peytavin G, Visseaux B, et al. Dolutegravir in HIV-2-Infected Patients With Resistant Virus to First-line Integrase Inhibitors From the French Named Patient Program. *Clin Infect Dis.* 2015;60:1521-7.
[\[PubMed Abstract\]](#) -
57. Pacheco P, Marques N, Rodrigues P, et al. Safety and Efficacy of Triple Therapy With Dolutegravir Plus 2 Nucleoside Reverse Transcriptase Inhibitors in Treatment-Naive Human Immunodeficiency Virus Type 2 Patients: Results From a 48-Week Phase 2 Study. *Clin Infect Dis.* 2023;77:740-8.
[\[PubMed Abstract\]](#) -
58. Gottlieb GS, Smith RA, Dia Badiane NM, et al. HIV-2 integrase variation in integrase inhibitor-naïve adults in Senegal, West Africa. *PLoS One.* 2011;6:e22204.
[\[PubMed Abstract\]](#) -
59. Smith RA, Raugi DN, Kiviat NB, et al. Phenotypic susceptibility of HIV-2 to raltegravir: integrase mutations Q148R and N155H confer raltegravir resistance. *AIDS.* 2011;25:2235-41.
[\[PubMed Abstract\]](#) -
60. Smith RA, Raugi DN, Pan C, et al. Three main mutational pathways in HIV-2 lead to high-level raltegravir and elvitegravir resistance: implications for emerging HIV-2 treatment regimens. *PLoS One.* 2012;7:e45372.
[\[PubMed Abstract\]](#) -
61. Le Hingrat Q, Collin G, Lê M, et al. A new mechanism of resistance of HIV-2 to integrase inhibitors: a 5 amino-acids insertion in the integrase C-terminal domain. *Clin Infect Dis.* 2019;4:657-67.
[\[PubMed Abstract\]](#) -
62. Raugi DN, Smith RA, Ba S, et al. Complex patterns of protease inhibitor resistance among antiretroviral treatment-experienced HIV-2 patients from Senegal: implications for second-line therapy. *Antimicrob Agents Chemother.* 2013;57:2751-60.
[\[PubMed Abstract\]](#) -
63. Desbois D, Roquebert B, Peytavin G, et al. In vitro phenotypic susceptibility of human immunodeficiency virus type 2 clinical isolates to protease inhibitors. *Antimicrob Agents Chemother.*

2008;52:1545-8.

[\[PubMed Abstract\]](#) -

64. Bénard A, Damond F, Campa P, et al. Good response to lopinavir/ritonavir-containing antiretroviral regimens in antiretroviral-naive HIV-2-infected patients. *AIDS*. 2009;23:1171-3.
[\[PubMed Abstract\]](#) -
65. Cavaco-Silva J, Aleixo MJ, Van Laethem K, et al. Mutations selected in HIV-2-infected patients failing a regimen including atazanavir. *J Antimicrob Chemother*. 2013;68:190-2.
[\[PubMed Abstract\]](#) -
66. Raugi DN, Smith RA, Gottlieb GS. Four Amino Acid Changes in HIV-2 Protease Confer Class-Wide Sensitivity to Protease Inhibitors. *J Virol*. 2016;90:1062-9.
[\[PubMed Abstract\]](#) -
67. Nowicka-Sans B, Gong YF, McAuliffe B, et al. In vitro antiviral characteristics of HIV-1 attachment inhibitor BMS-626529, the active component of the prodrug BMS-663068. *Antimicrob Agents Chemother*. 2012;56:3498-507.
[\[PubMed Abstract\]](#) -
68. Moranguinho I, Taveira N, Bárto I. Antiretroviral Treatment of HIV-2 Infection: Available Drugs, Resistance Pathways, and Promising New Compounds. *Int J Mol Sci*. 2023;24:5905.
[\[PubMed Abstract\]](#) -
69. Le Hingrat Q, Collin G, Bachelard A, et al. Ibalizumab shows in-vitro activity against group A and group B HIV-2 clinical isolates. *AIDS*. 2022;36:1055-60.
[\[PubMed Abstract\]](#) -
70. Borrego P, Calado R, Marcelino JM, et al. Baseline susceptibility of primary HIV-2 to entry inhibitors. *Antivir Ther*. 2012;17:565-70.
[\[PubMed Abstract\]](#) -
71. Borrego P, Taveira N. HIV-2 susceptibility to entry inhibitors. *AIDS Rev*. 2013;15:49-61.
[\[PubMed Abstract\]](#) -
72. Poveda E, Rodes B, Toro C, Soriano V. Are fusion inhibitors active against all HIV variants? *AIDS Res Hum Retroviruses*. 2004;20:347-8.
[\[PubMed Abstract\]](#) -
73. Smith RA, Raugi DN, Nixon RS, et al. Antiviral Activity of Lenacapavir Against HIV-2 Isolates and Drug-resistant HIV-2 Mutants. *J Infect Dis*. 2023 Dec 7. Online ahead of print.
[\[PubMed Abstract\]](#) -
74. Ba S, Raugi DN, Smith RA, et al. A Trial of a Single-tablet Regimen of Elvitegravir, Cobicistat, Emtricitabine, and Tenofovir Disoproxil Fumarate for the Initial Treatment of Human Immunodeficiency Virus Type 2 Infection in a Resource-limited Setting: 48-Week Results From Senegal, West Africa. *Clin Infect Dis*. 2018;67:1588-94.
[\[PubMed Abstract\]](#) -
75. Peterson K, Jallow S, Rowland-Jones SL, de Silva TI. Antiretroviral therapy for HIV-2 infection: recommendations for management in low-resource settings. *AIDS Res Treat*. 2011;2011:463704.
[\[PubMed Abstract\]](#) -
76. Matheron S, Damond F, Benard A, et al. CD4 cell recovery in treated HIV-2 infected adults is lower

than expected: results from the French ANRS CO5 HIV-2 cohort. *AIDS* 2006; 20:459-62.

[\[PubMed Abstract\]](#) -

77. Ekouevi DK, Tchounga BK, Coffie PA, et al. Antiretroviral therapy response among HIV-2 infected patients: a systematic review. *BMC Infect Dis*. 2014;14:461.
[\[PubMed Abstract\]](#) -
78. Drylewicz J, Matheron S, Lazaro E, et al. Comparison of viro-immunological marker changes between HIV-1 and HIV-2-infected patients in France. *AIDS*. 2008;22:457-68.
[\[PubMed Abstract\]](#) -
79. Charpentier C, Visseaux B, Bénard A, et al. Transmitted drug resistance in French HIV-2-infected patients. *AIDS*. 2013;27:1671-4.
[\[PubMed Abstract\]](#) -
80. Duarte F, Miranda AC, Peres S, et al. Transmitted drug resistance in drug-naïve HIV-2 infected patients. *AIDS*. 2016;30:1687-8.
[\[PubMed Abstract\]](#) -
81. Gilleece Y, Chadwick DR, Breuer J, et al. British HIV Association guidelines for antiretroviral treatment of HIV-2-positive individuals 2010. *HIV Med*. 2010;11:611-9.
[\[PubMed Abstract\]](#) -
82. Landman R, Damond F, Gerbe J, Brun-Vezinet F, Yeni P, Matheron S. Immunovirological and therapeutic follow-up of HIV-1/HIV-2-dually seropositive patients. *AIDS*. 2009;23:426-8.
[\[PubMed Abstract\]](#) -
83. Rodés B, Toro C, Jiménez V, Soriano V. Viral response to antiretroviral therapy in a patient coinfecting with HIV type 1 and type 2. *Clin Infect Dis*. 2005;41:e19-21.
[\[PubMed Abstract\]](#) -

References

- Álvarez M, Nevot M, Mendieta J, Martínez MA, Menéndez-Arias L. Amino acid residues in HIV-2 reverse transcriptase that restrict the development of nucleoside analogue resistance through the excision pathway. *J Biol Chem*. 2018;293:2247-2259.
[\[PubMed Abstract\]](#) -
- Bertine M, Gueudin M, Mélard A, et al. New Highly Sensitive Real-Time PCR Assay for HIV-2 Group A and Group B DNA Quantification. *J Clin Microbiol*. 2017;55:2850-2857.
[\[PubMed Abstract\]](#) -
- Branson BM, Mermin J. Establishing the diagnosis of HIV infection: new tests and a new algorithm for the United States. *J Clin Virol*. 2011;52 Suppl 1:S3-4.
[\[PubMed Abstract\]](#) -
- Brower ET, Bacha UM, Kawasaki Y, Freire E. Inhibition of HIV-2 protease by HIV-1 protease inhibitors in clinical use. *Chem Biol Drug Des*. 2008;71:298-305.
[\[PubMed Abstract\]](#) -
- Chang M, Steinmetzer K, Raugi DN, et al. Detection and differentiation of HIV-2 using the point-of-care Alere HIV-1/2 Detect nucleic acid test. *J Clin Virol*. 2017;97:22-25.
[\[PubMed Abstract\]](#) -

- Charpentier C, Camacho R, Ruelle J, et al. HIV-2EU: supporting standardized HIV-2 drug resistance interpretation in Europe. Clin Infect Dis. 2013;56:1654-8.
[\[PubMed Abstract\]](#) -
- Charpentier C, Eholié S, Anglaret X, et al. Genotypic resistance profiles of HIV-2-treated patients in West Africa. AIDS. 2014;28:1161-9.
[\[PubMed Abstract\]](#) -
- Charpentier C, Larrouy L, Collin G, et al. In-vitro phenotypic susceptibility of HIV-2 clinical isolates to the integrase inhibitor S/GSK1349572. AIDS. 2010;24:2753-5.
[\[PubMed Abstract\]](#) -
- de Silva TI, Leligdowicz A, Carlson J, et al. HLA-associated polymorphisms in the HIV-2 capsid highlight key differences between HIV-1 and HIV-2 immune adaptation. AIDS. 2018;32:709-714.
[\[PubMed Abstract\]](#) -
- Ekouevi DK, Balestre E, Coffie PA, et al. Characteristics of HIV-2 and HIV-1/HIV-2 Dually Seropositive Adults in West Africa Presenting for Care and Antiretroviral Therapy: The leDEA-West Africa HIV-2 Cohort Study. PLoS One. 2013;8:e66135.
[\[PubMed Abstract\]](#) -
- Esbjörnsson J, Månsson F, Kvist A, et al. Effect of HIV-2 infection on HIV-1 disease progression and mortality. AIDS. 2014;28:614-5.
[\[PubMed Abstract\]](#) -
- Gottlieb GS, Badiane NM, Hawes SE, et al. Emergence of multiclass drug-resistance in HIV-2 in antiretroviral-treated individuals in Senegal: implications for HIV-2 treatment in resource-limited West Africa. Clin Infect Dis. 2009;48:476-83.
[\[PubMed Abstract\]](#) -
- Gottlieb GS, Eholié SP, Nkengasong JN, et al. A call for randomized controlled trials of antiretroviral therapy for HIV-2 infection in West Africa. AIDS. 2008;22:2069-72.
[\[PubMed Abstract\]](#) -
- Gottlieb GS, Hawes SE, Kiviat NB, Sow PS. Differences in proviral DNA load between HIV-1-infected and HIV-2-infected patients. AIDS. 2008;22:1379-80.
[\[PubMed Abstract\]](#) -
- Greenberg AE. Possible protective effect of HIV-2 against incident HIV-1 infection: review of available epidemiological and in vitro data. AIDS. 2001;15:2319-21.
[\[PubMed Abstract\]](#) -
- Heitzinger K, Sow PS, Dia Badiane NM, et al. Trends of HIV-1, HIV-2 and dual infection in women attending outpatient clinics in Senegal, 1990-2009. Int J STD AIDS. 2012;23:710-6.
[\[PubMed Abstract\]](#) -
- Jallow S, Kaye S, Alabi A, et al. Virological and immunological response to Combivir and emergence of drug resistance mutations in a cohort of HIV-2 patients in The Gambia. AIDS. 2006;20:1455-8.
[\[PubMed Abstract\]](#) -
- Makvandi-Nejad S, Rowland-Jones S. How does the humoral response to HIV-2 infection differ from HIV-1 and can this explain the distinct natural history of infection with these two human retroviruses? Immunol Lett. 2015;163:69-75.

[[PubMed Abstract](#)] -

- Malloch L, Kadivar K, Putz J, et al. Comparative evaluation of the Bio-Rad Geenius HIV-1/2 Confirmatory Assay and the Bio-Rad Multispot HIV-1/2 Rapid Test as an alternative differentiation assay for CLSI M53 algorithm-I. J Clin Virol. 2013;58 Suppl 1:e85-91.
[[PubMed Abstract](#)] -
- Matheron S, Descamps D, Gallien S, et al. First-line Raltegravir/Emtricitabine/Tenofovir Combination in Human Immunodeficiency Virus Type 2 (HIV-2) Infection: A Phase 2, Noncomparative Trial (ANRS 159 HIV-2). Clin Infect Dis. 2018;67:1161-7.
[[PubMed Abstract](#)] -
- Miranda A, Peres S, Moneti V, Azevedo T, Aldir I, Mansinho K. Clinical and laboratorial impact of antiretroviral therapy in a cohort of Portuguese patients chronically infected with HIV-2. J Int AIDS Soc. 2014;17:19829.
[[PubMed Abstract](#)] -
- Owen SM, Ellenberger D, Rayfield M, et al. Genetically divergent strains of human immunodeficiency virus type 2 use multiple coreceptors for viral entry. J Virol. 1998;72:5425-32.
[[PubMed Abstract](#)] -
- Poulsen AG, Aaby P, Larsen O, et al. 9-year HIV-2-associated mortality in an urban community in Bissau, west Africa. Lancet. 1997;349:911-4.
[[PubMed Abstract](#)] -
- Raugi DN, Gottlieb GS, Sow PS, et al. HIV-1 outcompetes HIV-2 in dually infected Senegalese individuals with low CD4⁺ cell counts. AIDS. 2013;27:2441-50.
[[PubMed Abstract](#)] -
- Reeves JD, Doms RW. Human immunodeficiency virus type 2. J Gen Virol. 2002;83:1253-65.
[[PubMed Abstract](#)] -
- Rodés B, Sheldon J, Toro C, Jiménez V, Alvarez MA, Soriano V. Susceptibility to protease inhibitors in HIV-2 primary isolates from patients failing antiretroviral therapy. J Antimicrob Chemother. 2006;57:709-13.
[[PubMed Abstract](#)] -
- Roquebert B, Damond F, Collin G, et al. HIV-2 integrase gene polymorphism and phenotypic susceptibility of HIV-2 clinical isolates to the integrase inhibitors raltegravir and elvitegravir in vitro. J Antimicrob Chemother. 2008;62:914-20.
[[PubMed Abstract](#)] -
- Ruelle J, Roman F, Vandenbroucke AT, et al. Transmitted drug resistance, selection of resistance mutations and moderate antiretroviral efficacy in HIV-2: analysis of the HIV-2 Belgium and Luxembourg database. BMC Infect Dis. 2008;8:21.
[[PubMed Abstract](#)] -
- Smith RA, Raugi DN, Wu VH, et al. The Nucleoside Analog BMS-986001 Shows Greater In Vitro Activity against HIV-2 than against HIV-1. Antimicrob Agents Chemother. 2015;59:7437-46.
[[PubMed Abstract](#)] -
- Styer LM, Miller TT, Parker MM. Validation and clinical use of a sensitive HIV-2 viral load assay that uses a whole virus internal control. J Clin Virol. 2013;58 Suppl 1:e127-33.
[[PubMed Abstract](#)] -

- Thiébaud R, Matheron S, Taieb A, Brun-Vezinet F, Chêne G, Autran B. Long-term nonprogressors and elite controllers in the ANRS CO5 HIV-2 cohort. *AIDS*. 2011;25:865-7.
[\[PubMed Abstract\]](#) -
- Thomsen D, Erikstrup C, Jespersen S, et al. The influence of human leukocyte antigen-types on disease progression among HIV-2 infected patients in Guinea-Bissau. *AIDS*. 2018;32:721-728.
[\[PubMed Abstract\]](#) -
- Triki D, Cano Contreras ME, Flatters D, et al. Analysis of the HIV-2 protease's adaptation to various ligands: characterization of backbone asymmetry using a structural alphabet. *Sci Rep*. 2018;8:710.
[\[PubMed Abstract\]](#) -
- Tuaille E, Gueudin M, Lemée V, et al. Phenotypic susceptibility to nonnucleoside inhibitors of virion-associated reverse transcriptase from different HIV types and groups. *J Acquir Immune Defic Syndr*. 2004;37:1543-9.
[\[PubMed Abstract\]](#) -
- UNAIDS. Global AIDS Update 2023: the Path that Ends AIDS
[\[UNAIDS\]](#) -
- UNAIDS. HIV/AIDS. Data and statistics: 2022.
[\[UNAIDS\]](#) -
- Visseaux B, Charpentier C, Collin G, et al. Cenicriviroc, a Novel CCR5 (R5) and CCR2 Antagonist, Shows In Vitro Activity against R5 Tropic HIV-2 Clinical Isolates. *PLoS One*. 2015;10:e0134904.
[\[PubMed Abstract\]](#) -
- Visseaux B, Charpentier C, Hurtado-Nedelec M, et al. In vitro phenotypic susceptibility of HIV-2 clinical isolates to CCR5 inhibitors. *Antimicrob Agents Chemother*. 2012;56:137-9.
[\[PubMed Abstract\]](#) -
- Visseaux B, Charpentier C, Ozanne A, et al. Tropism distribution among antiretroviral-naive HIV-2-infected patients. *AIDS*. 2015;29:2209-12.
[\[PubMed Abstract\]](#) -
- Visseaux B, Charpentier C, Rouard C, et al. HIV-2 X4 tropism is associated with lower CD4+ cell count in treatment-experienced patients. *AIDS*. 2014;28:2160-2.
[\[PubMed Abstract\]](#) -
- Visseaux B, Damond F, Matheron S, Descamps D, Charpentier C. Hiv-2 molecular epidemiology. *Infect Genet Evol*. 2016;46:233-240.
[\[PubMed Abstract\]](#) -
- Wittkop L, Arsandaux J, Trevino A, et al. CD4 cell count response to first-line combination ART in HIV-2+ patients compared with HIV-1+ patients: a multinational, multicohort European study. *J Antimicrob Chemother*. 2017;72:2869-2878.
[\[PubMed Abstract\]](#) -
- Witvrouw M, Pannecouque C, Switzer WM, Folks TM, De Clercq E, Heneine W. Susceptibility of HIV-2, SIV and SHIV to various anti-HIV-1 compounds: implications for treatment and postexposure prophylaxis. *Antivir Ther*. 2004;9:57-65.
[\[PubMed Abstract\]](#) -

Figures

Figure 1 Characteristics of Persons Diagnosed with HIV-2, United States 2010-2017

Source: Peruski AH, Wesolowski LG, Delaney KP, et al. Trends in HIV-2 Diagnoses and Use of the HIV-1/HIV-2 Differentiation Test - United States, 2010-2017. MMWR Morb Mortal Wkly Rep. 2020;69:63-6.

This is a dynamic visualization. Please visit our website to experience this dynamic content.

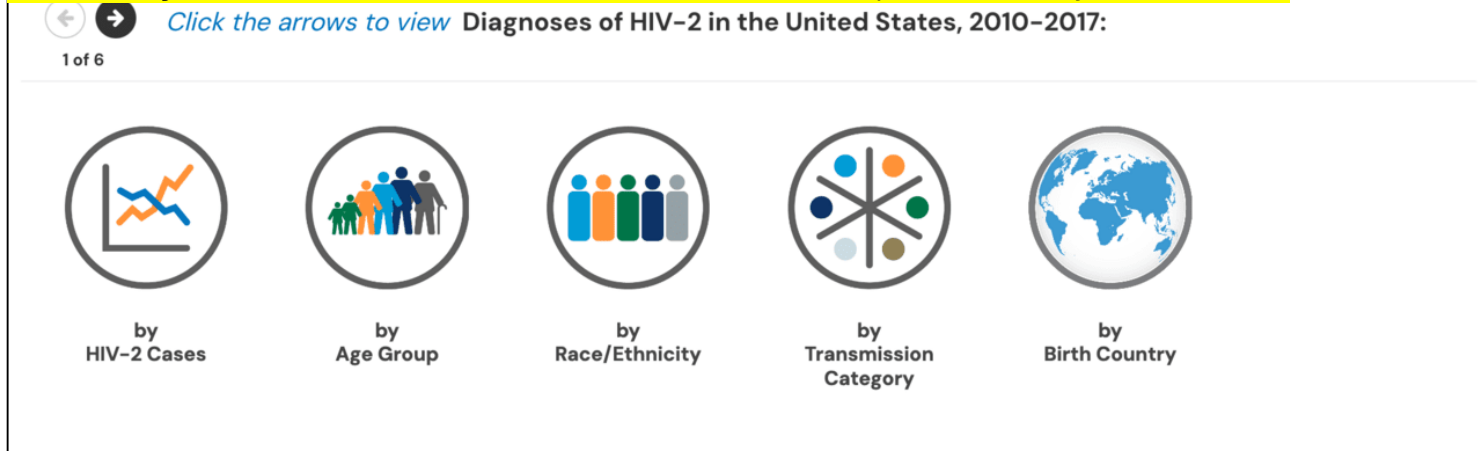


Figure 2 Median Time to AIDS and Death in Persons with HIV-1 or HIV-2 in West Africa

This graphic shows follow-up of 225 persons with HIV-1 and 87 with HIV-2. Median time to development of AIDS was slower in persons with HIV-2 but median survival was brief after AIDS in both groups.

Source: Esbjörnsson J, Månsson F, Kvist A, et al. Long-term follow-up of HIV-2-related AIDS and mortality in Guinea-Bissau: a prospective open cohort study. *Lancet HIV*. 2018;S2352-3018(18)30254-6.

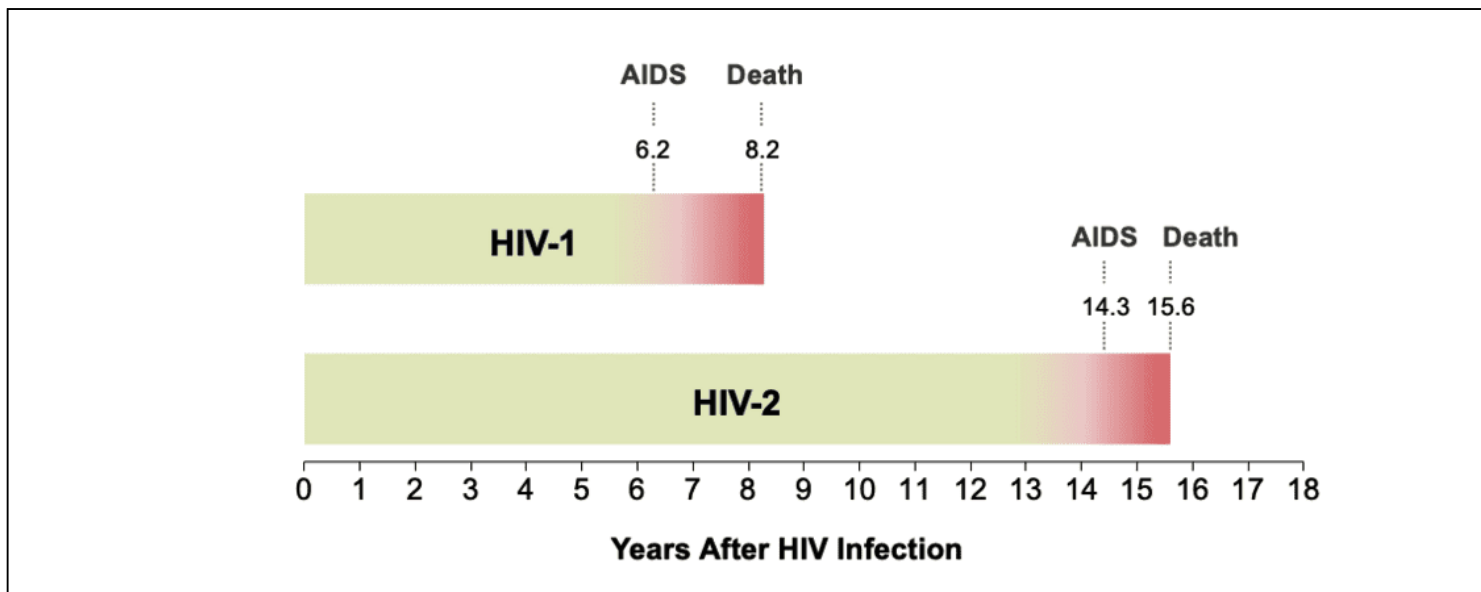


Figure 3 2018 CDC AHPL Recommended Laboratory HIV Testing Algorithm for Serum or Plasma Specimens

Abbreviation: APHL = Association of Public Health Laboratories

Source: Centers for Disease Control and Prevention and Association of Public Health Laboratories. 2018 Quick reference guide: Recommended laboratory HIV testing algorithm for serum or plasma specimens. Published January 27, 2018.

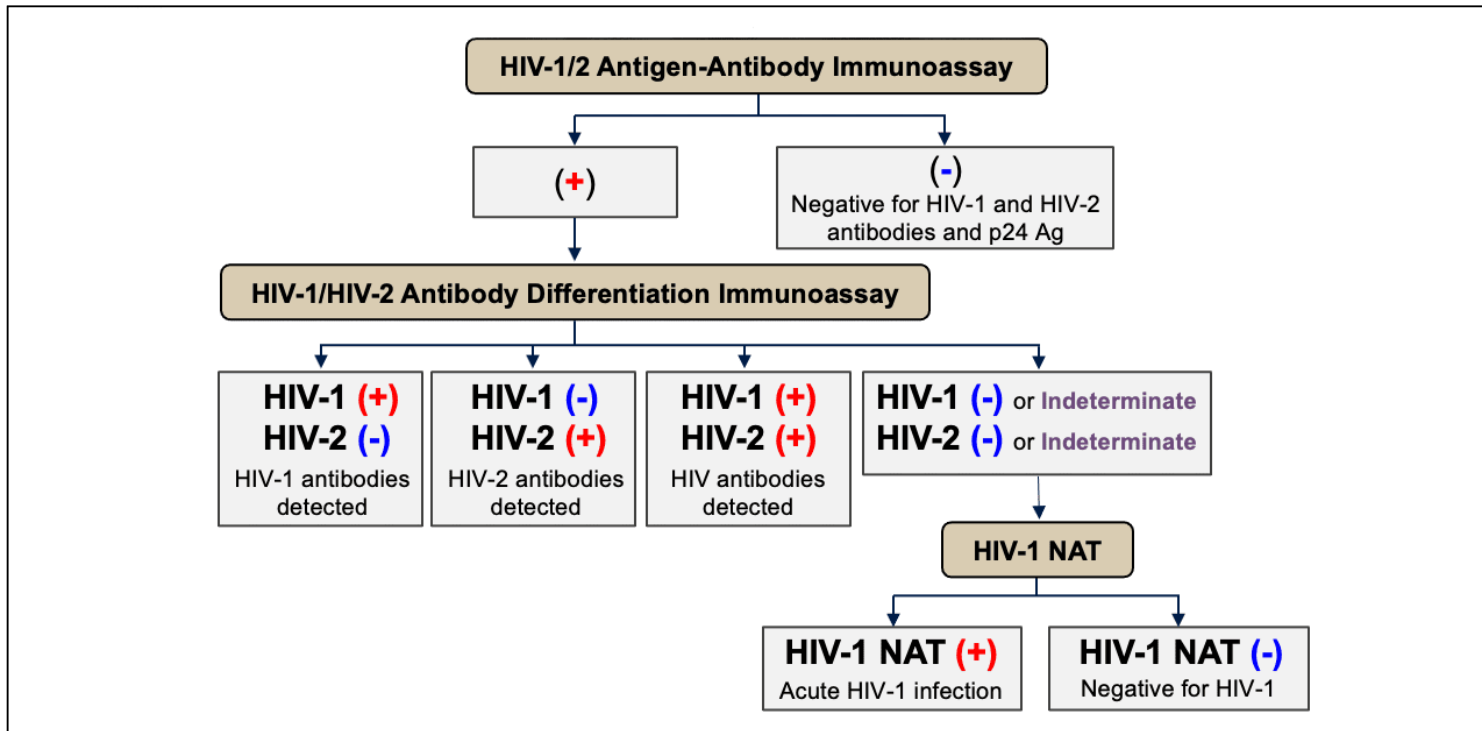


Figure 4 Geenius HIV-1/HIV-2 Supplemental Assay

The Geenius HIV-1/HIV-2 Supplemental Assay is a single-use immunochromatographic test that utilizes multiple recombinant or synthetic peptides to detect HIV-1 and HIV-2. Note the HIV-2 antibodies detected include gp36 and gp140 (marked by yellow color).

Source: modified from Fernández McPhee C, Álvarez P, Prieto L, et al. HIV-1 infection using dried blood spots can be confirmed by Bio-Rad Geenius™ HIV 1/2 confirmatory assay. J Clin Virol. 2015;63:66-9.

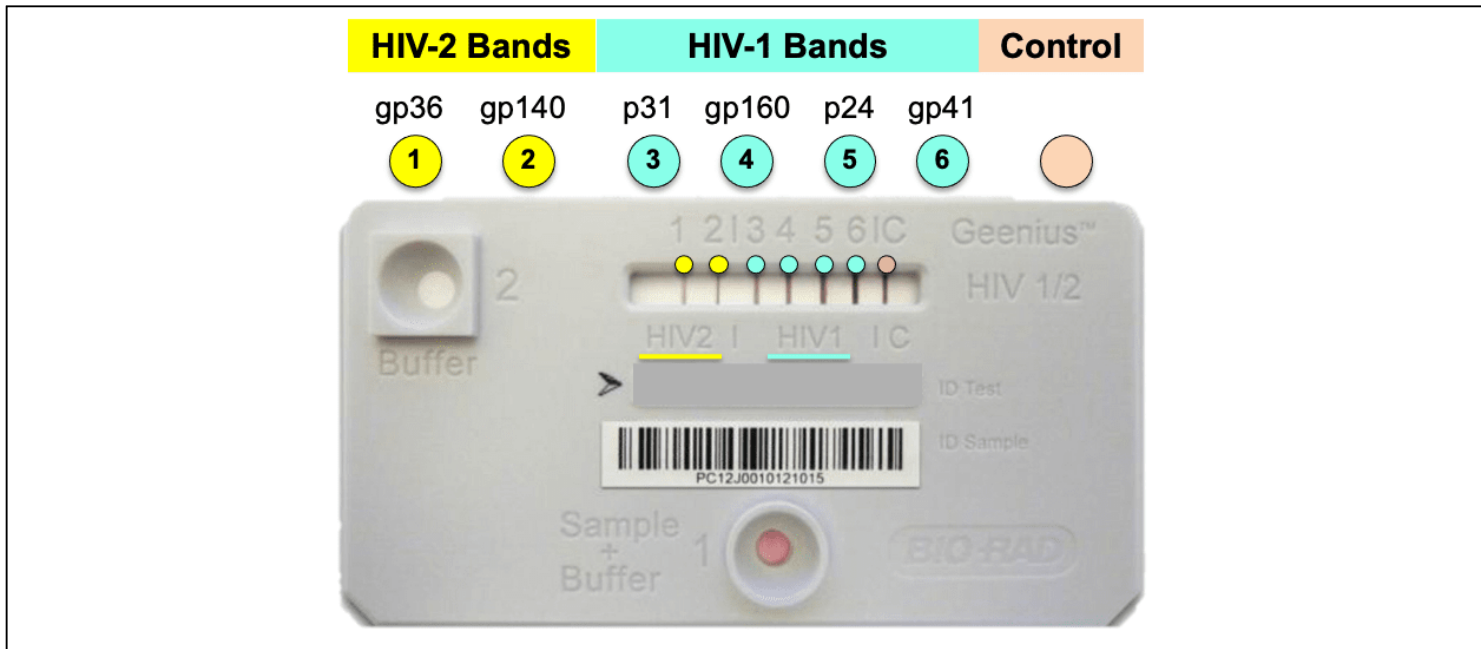


Figure 5 HIV-1 and HIV-2 Gene Products, Proteins, and Glycoproteins

Note the differences between some of the HIV-1 and HIV-2 proteins; this difference explains why HIV-1 Western blot tests fail to detect HIV-2 infection or give an indeterminate result.

Gene and Product	HIV-1	HIV-2
env		
Env Precursor	gp160	gp140
External Glycoprotein	gp120	gp105/125
Transmembrane Protein	gp41	gp36/41
pol		
Reverse Transcriptase	p66	p68
Reverse Transcriptase	p51	p53
Endonuclease	p31	p31/34
gag		
Gag Precursor	p55	p57
Matrix	p17	p17
Capsid	p24	p26
Nucleocapsid Precursor	p15	p15